## **CLAIMS**

## What is claimed is:

1. An engineered osteochondral graft for promoting the growth of cartilage in a patient at a defect site in need of repair, comprising a matrix block and a first population of MSCs, wherein said first population of MSCs are press-coated on a top surface of said matrix block, and said first population of MSCs forms a cartilage layer on said top surface of said matrix block.

10

- 2. The engineered osteochondral graft of **Claim 1**, wherein said matrix is biodegradable.
- 3. The engineered osteochondral graft of **Claim 2**, wherein said matrix is selected from the group consisting of demineralized bone matrix (DBM), biodegradable polymers, calcium-phosphates and hydroxyapatite.
- 4. The engineered osteochondral graft of **Claim 3**, wherein said matrix is a porous polylactic acid.
- 5. The engineered osteochondral graft of **Claim 4**, wherein said porous polylactic acid is D,D-L,L-polylactic acid.
- 6. The engineered osteochondral graft of **Claim 5**, wherein said matrix block is a D,D-L,L-polylactic acid polymer block of about 1 x 0.5 x 0.5 cm, said top surface of said matrix block is about 0.25 cm<sup>2</sup>, said first population of MSCs is about 1.5 x 10<sup>6</sup>, and said cartilage layer is about 1-1.5 mm thick.
- 7. The engineered osteochondral graft of Claim 1, wherein said matrix30 block has a shape compatible with said defect site.

5

10

- 8. The engineered osteochondral grafted of Claim 1, wherein said MSCs are isolated from a tissue selected from the group consisting of bone marrow, blood, periosteum, muscle, fat, bone and dermis.
- 9. The engineered osteochondral grafted of **Claim 8**, wherein said MSCs are isolated from bone marrow.
  - 10. The engineered osteochondral graft of **Claim 1**, wherein said engineered osteochondral graft further comprises an osteoinductive growth factor in an amount sufficient enough to elicit osseointegration.
  - 11. The engineered osteochondral graft of **Claim 10**, wherein said osteoinductive growth factor is BMP-2.
  - 12. The engineered osteochondral graft of **Claim 1**, wherein said engineered osteochondral graft further comprises a second population of MSCs which are loaded in the remaining volume of said matrix block, and said second population of MSCs is in an amount sufficient enough to elicit osseointegration.
- 13. The engineered osteochondral graft of **Claim 12**, wherein said engineered osteochondral graft further comprises an osteoinductive growth factor in an amount sufficient to elicit osseointegration.
- 14. The engineered osteochondral graft of **Claim 13**, wherein said osteoinductive growth factor is BMP-2.
  - 15. The engineered osteochondral graft of **Claim 1**, wherein said first population of MSCs are transiently or stably genetically engineered to express a gene product.

5

10

16. The engineered osteochondral graft of **Claim 15**, wherein said gene product is a member of the transforming growth factor-β superfamily.

- 17. A method of fabricating an osteochondral graft comprising the steps of contacting a top surface of a matrix block with a high-density pellet of a population of MSCs for a first period of time sufficient enough to form a cell-matrix structure, and culturing said cell-matrix structure in a chondrogenic differentiation medium for a second period of time sufficient enough to form a cartilage layer on said top surface of said matrix block, wherein said population of MSCs is an amount enough for the formation of said cartilage layer.
- 18. The method of **Claim 17**, wherein said chondrogenic differentiation medium contains a transforming growth factor.
- 19. The method of Claim 18, wherein said transforming growth factor is a member of TGF- $\beta$  superfamily
- 20. The method of **Claim 19**, wherein said member of TGF- $\beta$  superfamily is selected from the group consisting of TGF- $\beta$ 1, TGF- $\beta$ 3 and BMP-2.
- 21. The method of Claim 17, wherein said first population of MSCs is about  $1.5 \times 10^6$  cells per  $0.25 \text{ cm}^2$  of said top surface area.
- 25. The method of **Claim 17**, wherein said matrix block is a D,D-L,L-polylactic acid polymer block of about 1 x 0.5 x 0.5 cm, said top surface is about 0.25 cm², said population of MSCs is about 1.5 x 10<sup>6</sup>, said first period of time is about 3 hours, said second period of time is about 3 weeks, and said chondrogenic differentiation medium contains about 10 ng/ml TGF-β1.

5

10

- 23. A method of promoting the growth of cartilage in a patient at a site in need of repair, comprising the step of implanting an engineered osteochondral graft at said site, wherein said engineered osteochondral graft comprises a matrix block and a first population of MSCs, wherein said first population of MSCs are press-coated on a top surface of said matrix block, and said first population of MSCs forms a cartilage layer on said top surface of said matrix block.
- 24. The method of **Claim 23**, wherein said engineered osteochondral graft further comprises an osteoinductive growth factor in an amount sufficient enough to elicit osseointegration.
- 25. The method of **Claim 23**, wherein said engineered osteochondral graft further comprises a second population of MSCs which are loaded in the remaining volume of said matrix block, wherein said second population of MSCs is in an amount sufficient enough to elicit osseointegration.
- 26. The method of **Claim 25**, wherein said engineered osteochondral graft further comprises an osteoinductive growth factor in an amount sufficient to elicit osseointegration.
- 27. The method of **Claim 23**, wherein said first population of MSCs are transiently or stably genetically engineered to express a gene product.
- 25 28. The method of **Claim 27**, wherein said gene product is a member of the transforming growth factor-β superfamily.